

## **Lead (Pb) Deposition onto New and Biofilm-laden Potable Water Pipes**

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3

4 **Abstract**

5 Heavy metals' interactions with plumbing materials are complicated due to the differential formation  
6 of biofilms within pipes that can modulate, transform, and/or sequester heavy metals. This research  
7 aims to elucidate the mechanistic role of biofilm presence on Lead (Pb) accumulation onto  
8 crosslinked polyethylene (PEX-A), high-density polyethylene (HDPE), and copper potable water  
9 pipes. For this purpose, biofilms were grown on new pipes for three months. Five-day Pb exposure  
10 experiments were conducted to examine the kinetics of Pb accumulation onto the new and biofilm-  
11 laden pipes. Additionally, the influence of Pb initial concentration on the rate of its accumulation onto  
12 the pipes was examined. The results revealed greater biofilm biomass on the PEX-A pipes compared  
13 to the copper and HDPE pipes. More negative zeta potential was found for the biofilm-laden plastic  
14 pipes compared to the new plastic pipes. After five days of Pb exposure under stagnant conditions, the  
15 biofilm-laden PEX-A ( $980 \mu\text{g m}^{-2}$ ) and HDPE ( $1,170 \mu\text{g m}^{-2}$ ) pipe accumulated more than three times  
16 the Pb surface loading compared to the new PEX-A ( $265 \mu\text{g m}^{-2}$ ) and HDPE pipes ( $329 \mu\text{g m}^{-2}$ ),  
17 respectively. However, under flow conditions, Pb accumulation on biofilm-laden plastic pipes was  
18 lower than on the new pipes. Moreover, with increasing the initial Pb concentration, greater rates of  
19 Pb surface accumulation were found for the biofilm-laden pipes compared to the new pipes under  
20 stagnant conditions. First-order kinetics models best described the Pb accumulation onto both new  
21 and biofilm-laden water pipes under both stagnant and flow conditions.  
22

23 **Key Words:** Tap water quality, Plastic pipes, Lead, Biofilm, Heavy metals, Zeta potential  
24

25 **1. Introduction**

26 Drinking water quality can deteriorate substantially within building plumbing systems. The surface  
27 area to volume ratio of water pipes within buildings is significantly greater than that of water  
28 distribution systems, allowing greater physicochemical and microbiological interactions between the  
29 pipe wall and bulk water (Zlatanović et al., 2017). With the rapid movement toward sustainability,  
30 plastic pipes are increasingly being used to rehabilitate aging water infrastructure and construct new  
31 potable water systems, which reduces cost and ameliorates drinking water quality concerns associated  
32 with metal pipe corrosion. Plastic potable water pipes are corrosion-resistant and recyclable.  
33 Moreover, they have a lightweight, long service life and low carbon footprint (Hajibabaei et al.,  
34 2018). The global demand for plastic pipes has been estimated to grow by more than 4% per annum  
35 (Stewart, 2005). Cross-linked polyethylene (PEX) plumbing has emerged as a common material with

36 prevalent applications starting in the mid to late 1990s (Tech Topic: PEX, 2012; Currence, 2017).  
37 HDPE pipes initially replaced stormwater culverts in the 1980s and have been utilized for water  
38 supply pipes (Currence, 2017). Although plastic potable water pipes are significant alternatives to  
39 metallic pipes, the interactions between these pipes with microbial and heavy metal contaminants in  
40 the water remained largely unexplored. Research is needed to better understand the drivers of  
41 contaminant fate and transfer within these materials to ensure safe drinking water for consumers.

42         Lead (Pb) in tap water remains a serious ongoing threat to public health (DeSimone et al.,  
43 2020; Yang & Faust, 2019). Lead exposure can result in severe acute and chronic health impacts, such  
44 as irreversible developmental and behavioral delays in children (Jain et al., 2005; Edwards et al.,  
45 2009; Deshommes et al., 2016; Salehi et al., 2017; Rísová, 2019). Lead in tap water predominantly  
46 originates from the corrosion of lead-bearing plumbing materials, including lead service lines, brass  
47 valves and fittings, galvanized iron, lead-tin solder, and faucets (Boyd et al., 2008; Ghoochani et al.,  
48 2021). Lead service lines, which are frequently found in older buildings, can be connected to the  
49 recently renovated building plastic plumbing systems. Thus, service line corrosion products can be  
50 released into the building plumbing and subsequently accumulate on the inner surface of plastic pipes  
51 (Maas et al., 2007). Moreover, the brass fittings used for connecting the PEX plumbing materials  
52 could release heavy metals into the tap water. In our published field study, we quantified lead  
53 concentrations at cold and hot water fixtures during the first three months after installation of the new  
54 PEX plumbing. Even though new plastic plumbing was used, we found a greater level of lead inside  
55 the building than in the water collected from the service line (Salehi et al., 2018).

56         Despite the assumption by water utilities and regulatory agencies about the inert nature of  
57 plastic plumbing materials, our recent bench-scale experiments and field study demonstrated that  
58 plastic surfaces act as resting sites for heavy metals (Salehi et al., 2017; Salehi et al., 2018). Our  
59 recent studies (Huang et al., 2020; Salehi, 2022) and the findings of others (Wingender & Flemming,  
60 2004; Huang et al., 2017; Huang et al., 2019) showed that plastic pipes accumulate metal coatings in  
61 addition to biofilms on their surface after years of use. We demonstrated that the aged PEX-A ( $387 \mu\text{g m}^{-2}$ )  
62 and HDPE ( $418 \mu\text{g m}^{-2}$ ) pipes deposited significantly greater levels of Pb compared to the new  
63 PEX-A ( $288 \mu\text{g m}^{-2}$ ) and HDPE ( $335 \mu\text{g m}^{-2}$ ) pipes after 5 d of exposure to Pb aqueous solution of  
64  $300 \mu\text{g L}^{-1}$  under stagnant conditions (Huang et al., 2020). The research conducted by Huang et al.  
65 (2019) revealed that Pb was released by upstream brass and metal pipes onto the downstream PEX  
66 plumbing (Huang et al., 2019). Plastic pipe surfaces are generally exposed to Pb concentrations above  
67 the US Environmental Protection Agency (USEPA) action level ( $> 15 \mu\text{g L}^{-1}$ ). This exposure could be  
68 under stagnant water conditions, elevated temperatures, or pressurized conditions that might impact  
69 adsorption (Triantafyllidou & Edwards, 2012). Systematic research is needed to identify how heavy

70 metal uptake by plastic pipes differs from pellets, which have been studied extensively (Holmes et al.,  
71 2012, 2014; Huang et al., 2020), to better understand their fate in the built environment.

72 Biofilms are layers of amalgams of microorganisms and associated extracellular components  
73 that adhere to the surface of the pipe that is in direct contact with water (Kerr et al., 1998). They can  
74 interact with and influence metal ion sequestration within plumbing materials (Kerr et al., 1998;  
75 Costerton et al., 1987; Critchley et al., 2001). Although drinking water is disinfected prior to  
76 distribution, the disinfectant decay allows the survival of many microorganisms and their subsequent  
77 regrowth on the pipe's inner walls as biofilms. A recent study by Wang et al. (2022) revealed that  
78 plasticizers (e.g., phthalate ester) released by plastic potable water pipes into contact water promoted  
79 biofilm formation (Wang et al., 2022). Despite several studies conducted on biofilm accumulation on  
80 plastic potable water pipes in comparison to copper pipes, limited attention has been paid to the  
81 interaction of the biofilms present on the plastic pipe with heavy metals [e.g., Pb, Cu, Zn] that could  
82 be present in tap water. Our recent study revealed that the lead accumulated new and biofilm-laden  
83 plastic and copper potable water pipes could release their deposited lead as they were exposed to  
84 lower pH water under stagnant and flow conditions (Ghoochani et al., 2023).

85 Interactions between biofilms and scale may facilitate the continued development of both pipe  
86 surfaces. Reports suggest that water chemistry influences biofilm growth on plastic plumbing  
87 materials (Ji et al., 2015; Douterelo et al., 2016). While research has been conducted on the interactive  
88 effects of biofilms on pipe corrosion (Burleigh et al., 2014; Vargas et al., 2014; Rhoads et al., 2017;  
89 Marsili et al., 2018), the mechanistic role of biofilms present on plastic pipe surface on heavy metal  
90 sequestration has been overlooked. As our recent studies have revealed, the fundamental processes  
91 that control metal interactions with water infrastructure plastics remain poorly understood (Ahamed et  
92 al., 2020; Huang et al., 2020; Salehi, 2022). Thus, this research aims to elucidate the influence of  
93 biofilm presence on Pb deposition onto the PEX-A and HDPE potable water pipes compared to the  
94 copper pipes under water flow and stagnant conditions.

## 95 **2. Experimental**

### 96 **2.1 Materials**

97 Cross-linked polyethylene (PEX-A), high-density polyethylene (HDPE), and copper pipes were used  
98 in this study. The inner diameter of the PEX-A, HDPE, and copper pipes was 1.7 cm, 2.1 cm, and  
99 1.91 cm, respectively. The pipes were purchased from a local hardware store (Memphis, TN, USA).  
100 Inductively coupled plasma mass spectrometry (ICP-MS) lead (Pb) (1000 mg L<sup>-1</sup> in 3% nitric acid)  
101 standard solution was purchased from RICCA chemical company (Arlington, TX, USA). Clorox  
102 disinfectant comprised of 7.5% sodium hypochlorite (NaOCl) was purchased from a local store  
103 (Memphis, TN, USA). Nitric acid (70% purity) was purchased from Fisher Scientific (Hanover Park,

104 IL, USA). All the experiments were performed using ultrapure Milli-Q™ (18MΩ\*cm) treated water  
105 unless it is described otherwise.

## 106 **2.2 Pipe Disinfection and Biofilm Growth**

107 The PEX-A and HDPE pipe loops were constructed using ten pipes, each with a length of 2.7 m (9 ft),  
108 while for the copper pipe loop, ten pipes, each with a length of 3.0 m (10 ft) were used. Copper and  
109 PEX-A pipes were connected using brass connectors, while HDPE pipes were connected using  
110 polyvinyl chloride (PVC) connectors. After constructing the separate pipe loops, the pipes were shock  
111 disinfected, in which the pipe loops were filled with chlorinated municipal tap water ( $20 \pm 1 \text{ mg L}^{-1}$  as  
112  $\text{Cl}_2$ ) for 40 min. After that, each of the pipe loops was flushed with tap water for 30 min. Then, the  
113 pipe rigs were connected to the municipal tap water supply for biofilm development (**Figure SI-1a**).  
114 The municipal tap water was groundwater chlorinated for primary and secondary disinfection  
115 processes, aerated, and filtered. The corrosion inhibitor was orthophosphate ( $1 \text{ mg L}^{-1}$ ). Tap water ran  
116 through the pipe loops at  $2.0 \pm 0.2 \text{ L min}^{-1}$  for a daily cycle of 16 h of flow and 8 h of stagnation at  
117 room temperature to emulate standard household usage. Tap water quality parameters were measured  
118 weekly, including temperature, pH, dissolved oxygen (DO), and total chlorine concentrations. After  
119 biofilm generation for 90 d, pipes were removed from the stand and sectioned into 30 cm segments to  
120 undergo metal exposure experiments. In addition, representative pipe sections were used for microbial  
121 biomass and zeta potential measurements. Biofilm characterization was conducted using a droplet  
122 digital PCR system as described in **SI-1**.

## 123 **2.3 Zeta Potential Measurement**

124 Zeta potential measurements were conducted using an electrokinetic analyzer (SurPASS, 2010 model,  
125 Anton Paar USA, Ashland, VA) for new and biofilm-laden PEX-A and HDPE pipes to identify how  
126 the plastic pipe surface charge varies due to the biofilm accumulation. More information about  
127 mounting the samples and adjustment for analysis is described in **SI-2**. An automated titration was  
128 performed. After measuring the zeta potential at the original solution pH (usually between 5.5 and 6),  
129 small (~0.1 mL) doses of 0.1 N HCl were added to the solution until the pH dropped by at least 0.3  
130 pH units. The zeta potential was measured four times (two in one flow direction and two in the  
131 reverse flow direction) at each pH point. This continued until the solution pH was near 3. The solution  
132 was then replaced by 500 mL of fresh KCl (0.1 mM), and a similar titration was performed but with  
133 0.1 N NaOH until the pH was near 9. Total titration and measurement time were between 3 and 4 h.  
134 Data from the two runs were combined, and a titration curve of zeta potential versus pH between 3  
135 and 9 was generated.

## 136 **2.4 Lead (Pb) Exposure Experiments and Kinetics Modeling**

137 The synthetic tap water was used for the experiments to ensure that the chemical composition of the  
138 water used for the metal exposure experiments was consistent (Ahamed et al., 2020; Hadiuzzaman et  
139 al., 2023). A comparison of the chemical composition of the synthetic tap water and municipal water  
140 used for the biofilm growth process is shown in **Table SI-1**. The 5 d kinetics experiments were  
141 conducted in triplicate through six-time intervals (2, 6, 12, 24, 48, and 120 h) (Hadiuzzaman et al.,  
142 2022) under stagnant and flow conditions with initial Pb concentration of 300  $\mu\text{g L}^{-1}$ . To study the  
143 influence of initial Pb concentration on its accumulation onto the new and biofilm-laden water pipes,  
144 the pipe sections were exposed to five different aqueous solutions with Pb concentrations of 50, 150,  
145 500, 750, and 1000  $\mu\text{g L}^{-1}$  at pH=7.8 for 48 h. For the kinetics experiments that were conducted under  
146 flow conditions, 5.0 mL samples were collected from the water tank at different time intervals (2, 6,  
147 12, 24, 48, and 120 h). However, for studying the influence of Pb initial concentration, 5.0 mL  
148 samples were collected after 48 h. Additional information about Pb exposure experiments and control  
149 samples is provided in **SI-3**. Information regarding the water quality measurements and lead  
150 quantifications is provided in **SI-4**. The statistical analyses conducted to compare the Pb accumulation  
151 results are described in **SI-5**. The kinetics of Pb adsorption onto the new and biofilm-laden PEX-A,  
152 HDPE, and copper pipes were investigated through first-order and second-order reaction kinetics  
153 models where the equilibrium Pb surface loading was calculated by averaging the Pb surface loadings  
154 during the last two exposure periods (48 h and 120 h).

## 155 **3. Results And Discussions**

### 156 **3.1 Biomass Accumulation onto the Pipe Surfaces**

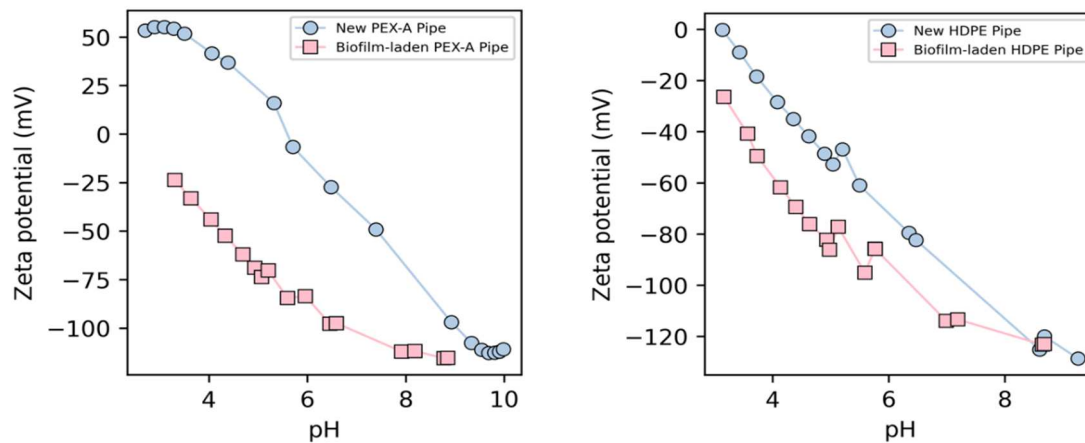
157 The biomass accumulation onto the inner pipe surfaces after three months of biofilm growth was  
158 studied through the quantification of 16S ribosomal RNA genes (rDNA) using triplicates. The results  
159 revealed the greatest ( $p$ -value < 0.05) initial biofilm formations were on PEX-A pipes ( $2.97 \times 10^9 \pm$   
160  $3.24 \times 10^8$  copies  $\text{cm}^{-2}$ , mean  $\pm$  standard deviation) compared to HDPE ( $1.19 \times 10^8 \pm 2.03 \times 10^8$  copies  
161  $\text{cm}^{-2}$ ) and copper pipes ( $1.07 \times 10^8 \pm 9.29 \times 10^7$  copies  $\text{cm}^{-2}$ ). The biofilm biomass was not statistically  
162 different among copper and HDPE pipes ( $p$ -value > 0.05). It is uncertain at this time if the  
163 communities that make up these biofilms are similar across this pipe material or if they are distinct,  
164 which may indicate differential microbial-mediated adsorption potential. Organics leached by plastic  
165 pipes [e.g., antioxidants, stabilizers, monomers] can further support the growth of microorganisms on  
166 the pipe surface (Salehi, et al., 2018; Salehi et al., 2020). The finding of a smaller extent of biofilm  
167 formation on the copper pipes versus the PEX-A pipes agrees with the other published studies that  
168 reported this result due to the antibacterial properties of the copper pipes (Inkinen et al., 2018). On  
169 the other hand, the organics leached by plastic pipes are reported that they promote microbial  
170 regrowth (Inkinen et al., 2018). PEX-A pipes are known to significantly contribute to the assimilable

171 organic carbon (AOC) release that could promote microbial regrowth within the pipes (Connell et al.,  
172 2016). The study conducted by Connell et al. (2016) revealed a greater rate of AOC release from PEX  
173 pipes compared to the HDPE pipes after 7 d exposure to the water (Connell et al., 2016). The  
174 quantification of dissolved organic carbon (DOC) release into the contact water from new HDPE and  
175 PEX-A pipes showed that  $1.1 \pm 0.3 \text{ mg L}^{-1}$  and  $1.9 \pm 0.1 \text{ mg L}^{-1}$  of DOC were leached into the contact  
176 water after 120 h of stagnation. The lower organics released by HDPE pipes compared to the PEX-A  
177 pipes could be the reason for a lower microbial growth on these pipes. Moreover, it should be noted  
178 that biofilm growth onto the PEX-A pipes was conducted from April to July, and biofilm growth onto  
179 the HDPE pipes was conducted from May to August. However, water flow and room temperature  
180 were constant during the biofilm growth process for all three types of pipes. The average temperature,  
181 total chlorine residuals, and DO concentration during the biofilm growth were recorded as  $21.2 \pm 1.2$   
182  $^{\circ}\text{C}$ ,  $0.9 \pm 0.1 \text{ mg L}^{-1}$ , and  $9.3 \pm 0.2 \text{ mg L}^{-1}$  for PEX-A pipes,  $22.7 \pm 1.2 \text{ }^{\circ}\text{C}$ ,  $1.0 \pm 0.1 \text{ mg L}^{-1}$ , and  $9.1 \pm$   
183  $0.2 \text{ mg L}^{-1}$  for HDPE pipes, and  $21.6 \pm 1.3 \text{ }^{\circ}\text{C}$ ,  $0.95 \pm 0.1 \text{ mg L}^{-1}$ , and  $9.3 \pm 0.1 \text{ mg L}^{-1}$  for copper  
184 pipes. Moreover, the microbial content and the physiochemical characteristics [e.g., temperature] of  
185 the water that entered the pipe loops might be slightly different and, consequently, may have  
186 influenced the extent of biomass accumulated onto each type of pipe.

### 187 **3.2 Surface Charge Variations Due to the Biofilm Accumulation**

188 Studying the zeta potential is the key parameter to understand how electrostatic attractions of lead  
189 species toward the pipe surface vary by biofilm presence. The literature indicated that the negative  
190 charge associated with the bacterial cells could promote electrostatic attractions of heavy metal ions  
191 present within the contact water and thus enhance their uptake (Wasserman et al., 2000). In this study,  
192 zeta potential measurements were conducted for new and biofilm-laden PEX-A and HDPE pipes to  
193 obtain a better insight into the pipes' surface chemistry, which influences their interactions with the  
194 charged Pb species present in the aqueous solution. As inferred from **Figure 1**, all new and biofilm-  
195 laden pipes demonstrated a negative surface charge at neutral pH. The extracellular polymer  
196 substances (EPS), lipoteichoic and lipopolysaccharide components present in biofilm tend to make  
197 more negative surface charge (Harper et al., 2019). The magnitude of zeta potential is reduced with  
198 increasing the pH. This finding corroborates those found by Chu et al. (2019) reported a negative zeta  
199 potential for microplastics (Chu et al., 2019). The zeta potential of both new and biofilm-laden pipes  
200 showed a similar descending trend with increasing the pH, with biofilm-laden PEX-A and HDPE  
201 pipes being slightly more negative than the new PEX-A and HDPE pipes. At low pH values, the  
202 hydrophilic sites on the pipe surface have a great tendency to adsorb protons due to the elevated  
203 proton concentration, which results in less negative zeta potential; however, with increasing the pH,  
204 the pipe surface becomes more negatively charged, this process called deprotonation (Liu, 2021).

205 The zeta potential of new and biofilm-laden PEX-A varied from 55 to -97 mV and -24 to -120  
 206 when increasing the pH from 3 to 8.9. The zeta potential of new and biofilm-laden HDPE varies  
 207 between 0 to -125 mV and -26 to -124 by increasing the pH from 3.1 to 8.6. The increased negativity  
 208 of zeta potential for the biofilm-laden pipes suggests that biofilms may facilitate general increases in  
 209 the magnitude of negative zeta potentials. Increasing the pipes' negative surface charge may result in  
 210 increased Pb cation attachment. This result is similar to the literature that indicated increasing in the  
 211 magnitude of negatively charged biofilms increases the likelihood of heavy metal sorption  
 212 (Kurniawan & Fukuda, 2022). Our recent studies demonstrated the water pH variation in PEX-A  
 213 potable water plumbing in a residential building to vary between 7.5 to 9.4 (Salehi et al., 2020).  
 214 Considering this pH range, it could be inferred that new and biofilm-laden plastic pipes have negative  
 215 surface charges. Additionally, the surface chemistry analysis of control samples that were prepared by  
 216 removal of biofilm from the biofilm-laden plastic pipes showed no surface oxidation. Thus, it could



217 be inferred that surface charge variation is solely due to biofilm growth (**Figure SI-2 and SI-3**).

218 (a)

(b)

219

220 **Figure 1.** The zeta potential variations versus pH for new and biofilm-laden (a) PEX-A and (b) HDPE  
 221 water pipe

222

### 223 3.3 Lead Accumulation onto the New and Biofilm-laden Water Pipes Under Stagnant Condition

224 In this study, the Pb accumulations onto the new and biofilm-laden PEX-A, HDPE, and copper pipes  
 225 under stagnant conditions and as a function of exposure duration were compared. As shown in **Figure**  
 226 **2**, during the entire exposure period, Pb accumulation onto the new copper pipes was significantly  
 227 greater than both new PEX-A ( $p$ -value < 0.05) and new HDPE pipes ( $p$ -value < 0.05), and even  
 228 biofilm-laden PEX-A ( $p$ -value < 0.05), and biofilm-laden HDPE pipes ( $p$ -value < 0.05). The  
 229 equilibrium Pb surface loading on new copper pipes ( $1,391 \mu\text{g m}^{-2}$ ) was more than 5 times greater  
 230 than new PEX-A ( $265 \mu\text{g m}^{-2}$ ) and 4 times greater than new HDPE pipes ( $329 \mu\text{g m}^{-2}$ ). However, the



231 biofilm presence significantly increased the Pb accumulation onto the PEX-A ( $p$ -value < 0.05) and  
232 HDPE ( $p$ -value < 0.05) pipes compared to their new pipes during the 5 d metal exposure period under  
233 stagnant conditions (**Figure 2**). At equilibrium, the biofilm-laden PEX-A pipes have more than three  
234 times greater Pb loading ( $980 \mu\text{g m}^{-2}$ ) compared to the new PEX-A pipes ( $265 \mu\text{g m}^{-2}$ ). It should be  
235 noted that our control experiments that examined the Pb loading in biofilm-laden PEX-A pipes before  
236 the metal exposure process showed that Pb surface loading on biofilm-laden PEX-A pipes was  $30.3$   
237  $\mu\text{g m}^{-2}$ , whereas it was below the detection limit for the new pipes prior to the metal exposure  
238 experiment.

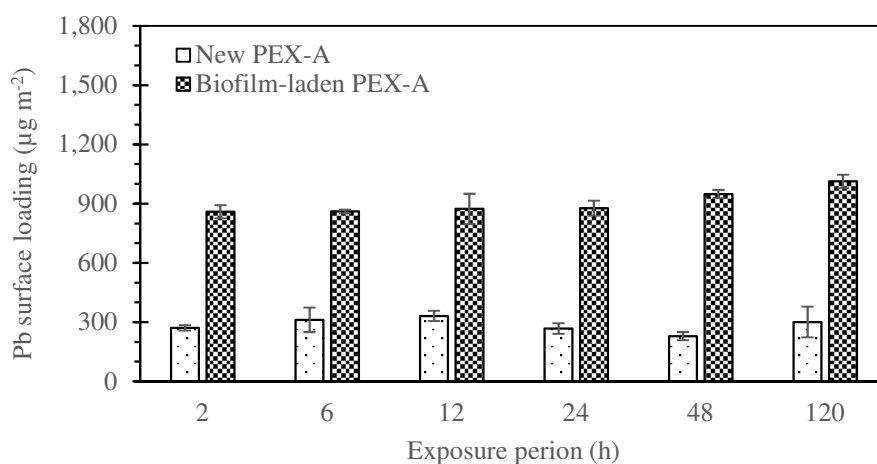
239 The biofilm-laden HDPE pipes accumulated approximately three times more Pb at  
240 equilibrium ( $1,170 \mu\text{g m}^{-2}$ ) than the new HDPE pipes ( $329 \mu\text{g m}^{-2}$ ). The biofilm-laden HDPE pipes  
241 accumulated 90% of its equilibrium Pb surface loading, while the new HDPE pipes obtained almost  
242 all their equilibrium Pb surface loadings during the first two hours of Pb exposure. The biofilm-laden  
243 HDPE pipes acquired 85% of  $[\text{Pb}]_i$  in the synthetic tap water as their equilibrium surface loading after  
244 5 d of metal exposure experiments; however, the new HDPE pipes only accumulated 24% of  $[\text{Pb}]_i$  at  
245 equilibrium. It should be noted that our control experiments that examined the Pb loading in biofilm-  
246 laden HDPE pipes before the metal exposure process showed that Pb surface loading on biofilm-laden  
247 HDPE pipes was  $44.3 \mu\text{g m}^{-2}$ , whereas it was below the detection limit for the new pipes prior to the  
248 metal exposure experiment. The 1 ft HDPE pipe segment was exposed to a greater volume of Pb  
249 aqueous solution (92 mL) compared to the 1 ft PEX-A pipe segment (65 mL) because of its larger  
250 diameter than PEX-A pipes. The initial Pb mass to surface area ratio for a PEX-A pipe segment was  
251  $1.2 \text{ mg m}^{-2}$  however it was  $1.4 \text{ mg m}^{-2}$  for an HDPE pipe segment. Thus, a greater mass of Pb was  
252 available within the HDPE pipe system, which may have contributed to a greater Pb surface loading  
253 on both new and biofilm-laden HDPE pipes. Moreover, a different rate of organic leaching by plastic  
254 pipes may have influenced the Pb uptake behavior (Kelley et al., 2014; Salehi et al., 2020). The  
255 reduction of Pb surface loading on biofilm-laden HDPE pipes after 2 h of exposure, could be due to  
256 the release of initially accumulated Pb species that were loosely attached to the biofilm surface back  
257 into the water. However, an overall increasing rate of Pb accumulation for biofilm-laden HDPE pipes  
258 was found over the exposure duration of 5 d.

259 Despite the findings for PEX-A and HDPE pipes, the biofilm accumulation onto the copper  
260 pipes did not significantly increase the Pb accumulation under stagnant conditions. The biofilm-laden  
261 copper pipes ( $1,527 \mu\text{g m}^{-2}$ ) accumulated more than the new copper pipe ( $1,352 \mu\text{g m}^{-2}$ ), whereas the  
262 difference was not statistically significant ( $p$ -value > 0.05). At the equilibrium 93% of total Pb in the  
263 synthetic tap water was accumulated on the biofilm-laden copper pipe and 85% of total Pb was  
264 accumulated on new copper pipes. For copper pipes, biofilm surface accumulation is known to  
265 promote pipe corrosion and consequently release copper ions. Some of these released copper ions

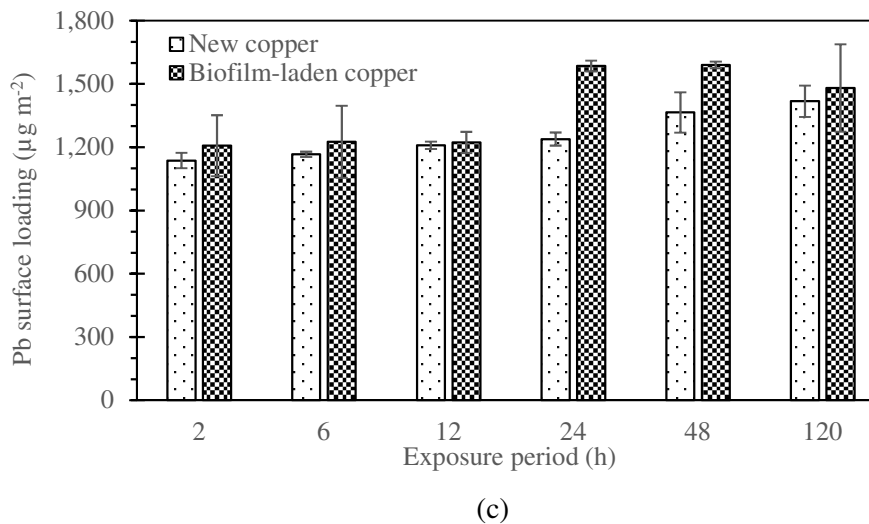
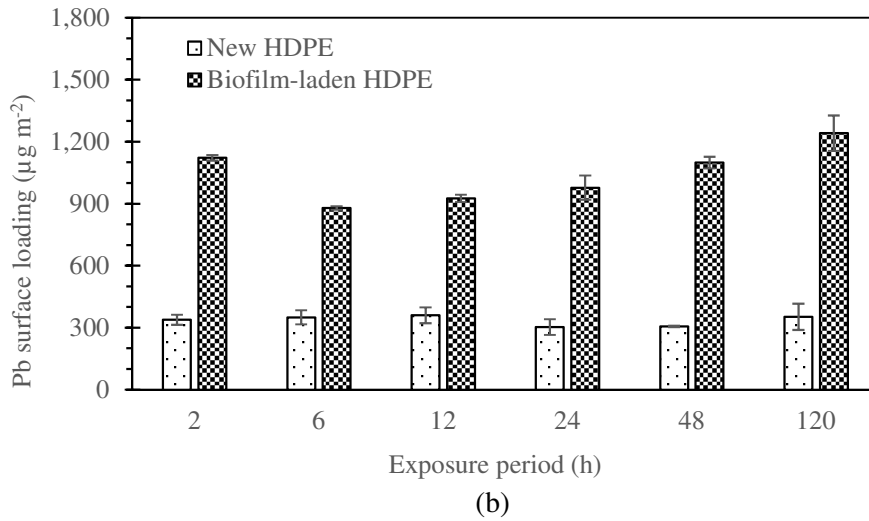
266 could be stored within the biofilm structure, forming the complexation with the ligands that are  
267 already present within the extracellular polymeric substance of the biofilm (Galarce et al., 2020). The  
268 occupation of these available surface sites by  $\text{Cu}^{2+}$  ions likely reduce the available sites to accumulate  
269  $\text{Pb}^{2+}$  species during the metal exposure experiments.

270 The ligands present within our synthetic tap water [ $\text{OH}^-$ ,  $\text{SO}_4^{2-}$ ,  $\text{NO}_3^-$ ,  $\text{Cl}^-$ ,  $\text{HPO}_4^{2-}$ ,  $\text{HCO}_3^-$ ,  
271  $\text{SiO}_3^{2-}$ ] could form both dissolved [e.g.,  $\text{Pb}^{2+}$ ,  $\text{Pb}(\text{OH})^+$ ] and insoluble Pb [e.g.,  $\text{Pb}(\text{OH})_2$ ,  $\text{Pb}_3(\text{PO}_4)_2$ ]  
272 complexes with  $\text{Pb}^{2+}$ . The Pb exposure experiments were conducted at the initial concentration of  
273  $[\text{Pb}]_i=300 \mu\text{g L}^{-1}$  and  $\text{pH}=7.8$ . However, the aqueous system was closed, pH may have been changed  
274 as we conducted the metal exposure experiments. But we expect to have mostly Pb precipitates to be  
275 present in the system. The  $K_s$  values for  $\text{Pb}(\text{OH})_2$ ,  $\text{PbCO}_3$ , and  $\text{Pb}_3(\text{PO}_4)_2$  are  $4.2 \times 10^{-15}$ ,  $1.5 \times 10^{-13}$ ,  
276 and  $1.0 \times 10^{-32}$  (SenGupta, 2017). So,  $\text{Pb}_3(\text{PO}_4)_2$  controls the aqueous Pb concentration in this system.  
277 Thus, both adsorption and precipitation processes [accumulation] have occurred simultaneously in the  
278 studied aqueous system. As we reported earlier, the metal species adsorbed onto the plastic surface  
279 could act as nucleation sites to grow the precipitates. The low energy polymer sites, such as surface  
280 impurities, polymer terminal groups, or an arrangement of polymeric chains, could act as the  
281 nucleation sites for the precipitate formation (Salehi et al., 2017).

282



(a)



283  
 284 **Figure 2.** The results for Pb surface loadings on new and biofilm-laden (a) PEX-A ( $p$ -value < 0.05),  
 285 (b) HDPE ( $p$ -value < 0.05), and (c) Copper ( $p$ -value > 0.05) water pipes over time, under stagnant  
 286 condition  
 287

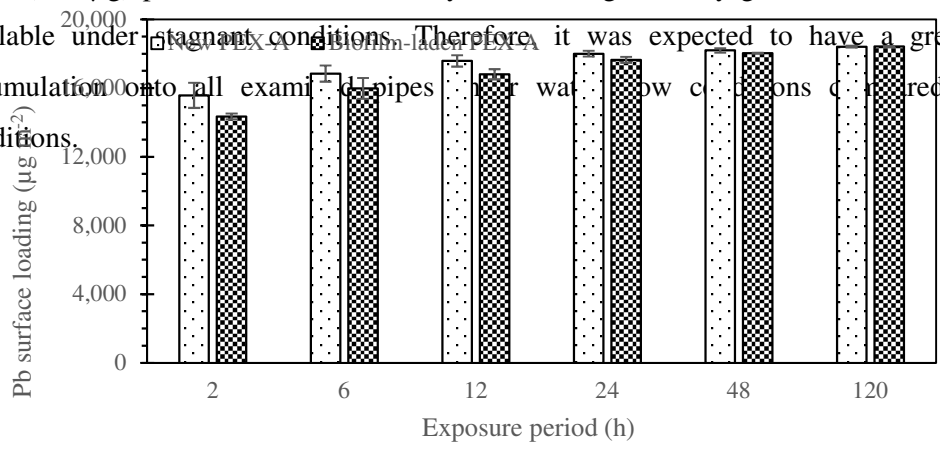
### 288 3.4 Lead Accumulation onto the New and Biofilm-laden Water Pipes Under Flow Condition

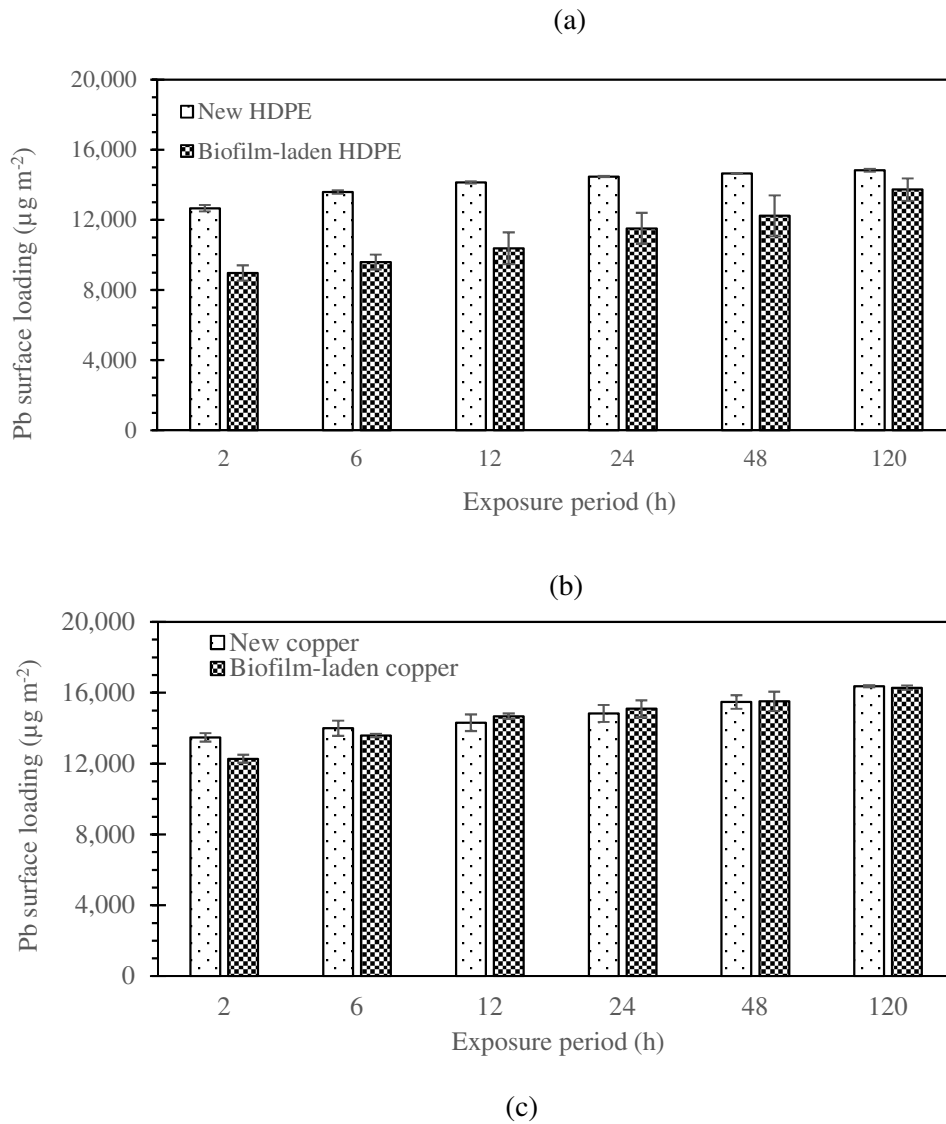
289 The Pb accumulation onto the plastic and copper water pipes under flow conditions was compared  
 290 with the stagnant condition to better understand how variations of water use behavior within the  
 291 residential and commercial buildings influence the extent of Pb accumulation onto the potable water  
 292 pipes. Unlike our findings of increased Pb accumulation onto the plastic pipes due to the biofilm  
 293 presence under stagnant conditions, we have found significant reductions ( $p$ -value < 0.05) in Pb  
 294 surface loadings on both biofilm-laden PEX-A ( $p$ -value < 0.05) and HDPE ( $p$ -value < 0.05) pipes  
 295 compared with their new pipes, under water flow conditions (**Figure 3**). This might be due to the  
 296 shorter duration of Pb species interactions with the biofilm surface under flow conditions, which may  
 297 not be sufficient to allow their removal from the aqueous system. The metal exposure experiments  
 298 were conducted under consistent flow conditions to prevent the creation of the shear forces and

299 subsequent biofilm detachment. Although, it might be possible that some of the metal accumulated  
 300 biofilms were released into the water due to their reduced internal cohesive strength caused by their  
 301 aging or its nutrient starvation as exposed to the synthetic water and not the tap water (Boe-Hansen et  
 302 al., 2002; Abe et al., 2012). **Table SI-1** demonstrates the differences in the chemical composition of  
 303 synthetic tap water used for Pb exposure experiments and the municipal tap water that was used for  
 304 the three months of the biofilm growth process.

305 Pb surface loading on the new copper pipes ( $15,918 \mu\text{g m}^{-2}$ ) was not significantly different  
 306 from on their biofilm-laden pipes ( $15,898 \mu\text{g m}^{-2}$ ) ( $p$ -value  $> 0.05$ ). The equilibrium Pb surface  
 307 loadings onto the new and biofilm-laden PEX-A pipes were found as  $18,300 \mu\text{g m}^{-2}$  and  $18,219 \mu\text{g m}^{-2}$ ,  
 308 significantly greater ( $p$ -value  $< 0.05$ ) than new ( $14,739 \mu\text{g m}^{-2}$ ) and biofilm-laden HDPE ( $12,938$   
 309  $\mu\text{g m}^{-2}$ ) pipes, respectively. Under flow conditions, a similar volume of 1 L was circulated in both  
 310 PEX and HDPE pipe loops. Thus, available Pb mass was similar in both systems. However, a smaller  
 311 surface area was exposed to the Pb aqueous solution in the PEX pipe segment ( $0.016 \text{ m}^2$ ) compared to  
 312 the HDPE pipe segment ( $0.02 \text{ m}^2$ ). This might have contributed to the PEX-A pipes' greater Pb  
 313 surface loading compared to the HDPE pipes under the flow conditions. Moreover, due to the greater  
 314 volume of the Pb aqueous solution, the influence of organics leached by plastic pipes might be smaller  
 315 compared to the stagnant condition. It should be mentioned that during our water flow experiments, a  
 316 total of 1 L Pb aqueous solution of  $[\text{Pb}]_i=300 \mu\text{g L}^{-1}$  at  $\text{pH}=7.8$  was circulated through the 30 cm pipe  
 317 sections; however, in our stagnant experiments, the PEX-A, HDPE, and copper pipe sections were  
 318 filled with 65 mL, 92 mL, and 100 mL of the Pb aqueous solution, respectively. Thus, the total mass  
 319 of Pb ( $300 \mu\text{g}$ ) present within the flow system was significantly greater than the Pb mass that was  
 320 available under stagnant conditions. Therefore, it was expected to have a greater level of Pb

321 accumulation onto all examined pipes for water flow conditions compared to the stagnant  
 322 conditions.





323 **Figure 3.** The results for Pb surface loadings on new and biofilm-laden (a) PEX-A ( $p$ -value < 0.05),  
 324 (b) HDPE ( $p$ -value < 0.05) and (c) Copper ( $p$ -value > 0.05) water pipes over time under flow  
 325 condition

326

327 **3.5 The Kinetics Modeling for Lead Accumulation onto the New and Biofilm-Laden Water**  
 328 **Pipes Under Stagnant and Flow Conditions**

329 In this study, kinetics modeling was conducted to provide a better mechanistic understanding of the  
 330 combined physiochemical mechanisms that control the rate of Pb accumulation onto the new and  
 331 biofilm-laden potable plastic and copper water pipes. For this purpose, first-order and second-order  
 332 reaction kinetics models were applied. The first-order kinetics model assumes that the rate of changes  
 333 in the solute uptake over time is proportional to the difference in the saturation concentration and the  
 334 amount of adsorbed solute over time (Romanov et al., 1998; Sahoo & Prelot, 2020). The first order  
 335 reaction model mostly fits the experimental metal adsorption data when the solute adsorption  
 336 proceeds through the diffusion to the solid micropores. However, the second-order reaction model  
 337 assumes that the rate-limiting step is the chemisorption process (López-Luna et al., 2019; Sahoo &

338 Prelot, 2020). A non-linear chi-square ( $\chi^2$ ) comparison tests were conducted between the models (first  
 339 and second order) to determine the best-fitted kinetics model for the studied pipes (**Table 1**).

340 The lower  $\chi^2$  values found here suggest that the first-order kinetics model better describes the  
 341 Pb accumulation onto almost all water pipes tested. Thus, the diffusion of Pb species from the bulk  
 342 aqueous phase to the pipe surface was likely the primary mechanism of Pb accumulation onto the pipe  
 343 surface. This agrees with the Azizian (2004) theoretical study which demonstrated that at the high  
 344 initial adsorbate concentration, the adsorption process of the kinetics studies can be described by the  
 345 first-order reaction model, however, at low initial adsorbate concentration, it obeys the second-order  
 346 reaction model (Azizian, 2004). The half-life ( $t_{1/2}$ ) calculation revealed a longer duration half-life for  
 347 biofilm-laden PEX-A (6.7 h) and HDPE (9.9 h) pipes compared to the new PEX-A (0.4 h) and HDPE  
 348 (0.6 h) pipes. This finding suggests that Pb accumulation onto the new plastic pipes occurs more  
 349 rapidly, as the most readily available surface deposition sites were rapidly occupied by the Pb species.  
 350 However, despite the greater adsorption capacity of the biofilm-laden plastic pipes, they were not  
 351 readily accessible for the Pb species, and consequently, a longer duration took to reach the half-life. It  
 352 was completely different for the copper pipe as the half-life for new copper pipes (6.9 h) was greater  
 353 than the half-life for the biofilm-laden copper pipe (1.8 h).

354 The kinetics data collected under water flow conditions also revealed that first-order reaction  
 355 models best describe the Pb accumulation onto the new and biofilm-laden water pipes. This is clearly  
 356 a mass transfer process due to the abundance of available Pb species in the system as water circulates  
 357 through the pipes. The increased half-lives for the plastic pipes and decreased half-lives for the metal  
 358 pipes due to the presence of biofilm were similar under both stagnant and flow conditions; however,  
 359 the difference in half-lives between the new and biofilm-laden pipes under flow conditions was  
 360 smaller than the one's under stagnant conditions. It can be due to the abundance of Pb species in the  
 361 system as water circulated in the pipes. Moreover, this finding explains our results regarding the lower  
 362 Pb accumulation onto the biofilm-laden plastic pipes compared to the new plastic pipes under flow  
 363 conditions due to the short durations of Pb species contact with the biofilm that was not sufficient to  
 364 allow a significant uptake of Pb species. Moreover, the convective diffusion in the flow conditions  
 365 may have resulted in a faster mass transfer than pure diffusion occurred under stagnant conditions.

366 **Table 1.** The kinetics models' parameters and non-linear chi-square ( $\chi^2$ ) values for Pb accumulation  
 367 onto the new and biofilm-laden water pipes under stagnant and flow conditions

Pipe		Model	First-order reaction model					Second-order reaction model		
			$q_{e, \text{exp}}$ ( $\mu\text{g m}^{-2}$ )	$q_{e, \text{mod}}$ ( $\mu\text{g m}^{-2}$ )	$k_1$ ( $\text{h}^{-1}$ )	$t_{1/2}$ (h)	$\chi^2$	$q_{e, \text{mol}}$ ( $\mu\text{g m}^{-2}$ )	$k_2 \times 10^3$ ( $\text{m}^2 \mu\text{g}^{-1} \text{h}^{-1}$ )	$\chi^2$
Stagnant	New	Copper	1,391	1,416	0.101	6.9	0.42	1450	0.244	2.38
		PEX-A	265	288	1.890	0.4	1.87	285	90.0	1.48
		HDPE	329	335	1.200	0.6	0.12	356	3.0	2.12

Flow	Biofilm-laden	Copper	1,535	1,553	0.381	1.8	0.18	1496	1.01	1.02	
		PEX-A	981	1,011	0.104	6.7	0.91	1035	0.305	2.93	
		HDPE	1,170	1,275	0.070	9.9	8.61	1286	0.183	10.49	
	New	Biofilm-laden	Copper	15,917	16,380	0.090	7.7	13.08	16,769	0.02	43.21
			PEX-A	18,300	18,410	0.148	4.7	0.66	18,535	0.066	2.97
			HDPE	14,739	14,835	0.147	4.7	0.61	14,491	0.077	2.73
		New	Copper	15,898	16,284	0.107	6.5	9.16	16,648	0.022	33.81
			PEX-A	18,219	18,412	0.122	5.7	2.03	18,643	0.036	9.65
			HDPE	12,938	13,734	0.073	9.6	41.03	14,501	0.01	158.80

368

### 369 **3.6 Influence of Lead (Pb) Initial Concentration on the Rate of its Accumulation onto the Water** 370 **Pipes**

371 The influence of Pb initial concentration on the rate of its accumulation onto the new and biofilm-  
372 laden water pipes was investigated under stagnant (**Figure 4a**) and water flow conditions (**Figure 4b**).  
373 Under stagnant conditions, both new and biofilm-laden water pipes revealed a similar order of copper  
374 > PEX-A > HDPE for the increased rate of Pb surface accumulation by increasing the Pb initial  
375 concentration, and the accumulation onto the pipes was significantly different ( $p$ -value < 0.05). Under  
376 stagnant conditions, biofilm presence drastically increased the rate of Pb accumulation onto all water  
377 pipes with increasing the Pb concentration. A linearly increased Pb surface loading was found for new  
378 and biofilm-laden PEX-A and HDPE pipes ( $R^2 \geq 0.90$ ) and copper pipes ( $R^2 \geq 0.84$ ) with increasing  
379 the Pb concentration under stagnant conditions. By applying a linear model, the slopes are defined as  
380 the Pb accumulation rate at each condition. The Pb accumulation rate for new PEX-A, HDPE, and  
381 copper pipes under stagnant conditions varied from 0.004, 0.003, and 0.002 m to 0.099, 0.014, and  
382 0.011 m due to the presence of biofilm.

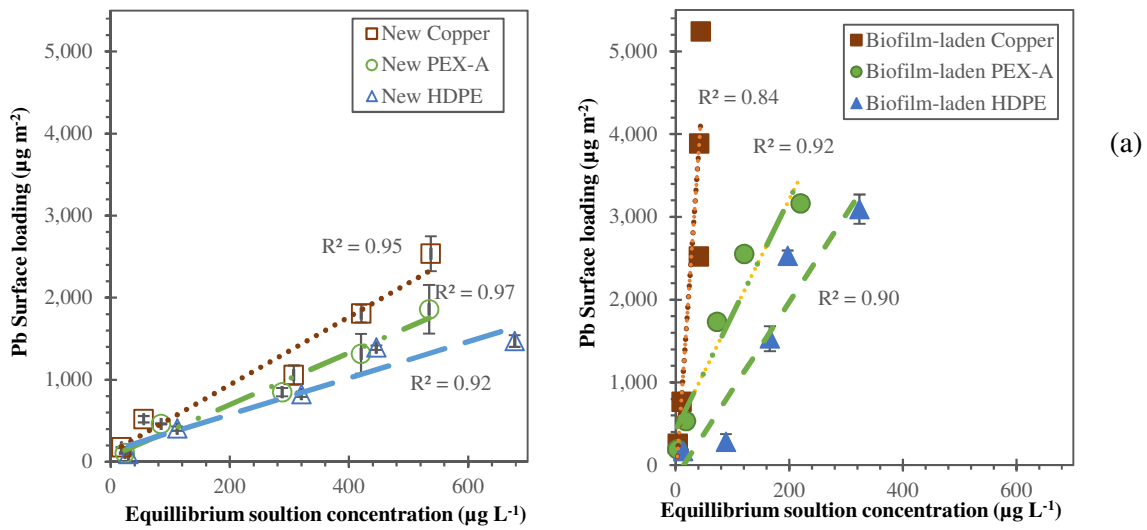
383 Under stagnant condition, the greatest Pb surface loading onto the new PEX-A, HDPE, and  
384 copper pipes were found as 1,859, 1,472, and 2,536  $\mu\text{g m}^{-2}$  and were significantly smaller than the  
385 maximum average Pb surface loadings onto the biofilm-laden PEX-A (3,162  $\mu\text{g m}^{-2}$ ), HDPE (3,094  
386  $\mu\text{g m}^{-2}$ ), and copper pipes (5,240  $\mu\text{g m}^{-2}$ ), respectively. Under flow conditions, the Pb surface loading  
387 onto the new PEX-A, HDPE, and copper pipes increased linearly with raising the Pb concentration  
388 ( $R^2 \geq 0.84$ ) (**Figure 4b**). Despite a similar trend, the coefficient of determination for this linear  
389 regression reduced when biofilm was present on the pipe surfaces ( $R^2 \leq 0.74$ ) (**Figure 4b**). The  
390 greatest Pb surface loadings onto the new PEX-A, HDPE, and copper pipes under flow conditions  
391 varied from 58,427, 46,947, and 48,579  $\mu\text{g m}^{-2}$  to 51,468, 29,185, and 41,288  $\mu\text{g m}^{-2}$  due to biofilm  
392 presence. At almost all concentrations, less Pb surface loadings were found on the biofilm-laden water  
393 pipes compared to the new pipes under flow conditions.

394 The Pb accumulation rates onto the new PEX-A (120.49 m), HDPE (248.5 m), and copper  
395 pipes (0.688 m) reduced to the biofilm-laden PEX-A (0.397 m), HDPE (0.082 m), and copper pipes  
396 (0.234 m). As it was mentioned earlier, the short contact of the Pb species and biofilm surface during  
397 the water flow experiments may have resulted in insufficient interaction between those and,  
398 consequently, a lower accumulation rate compared to the new plastic pipes. Moreover, although the

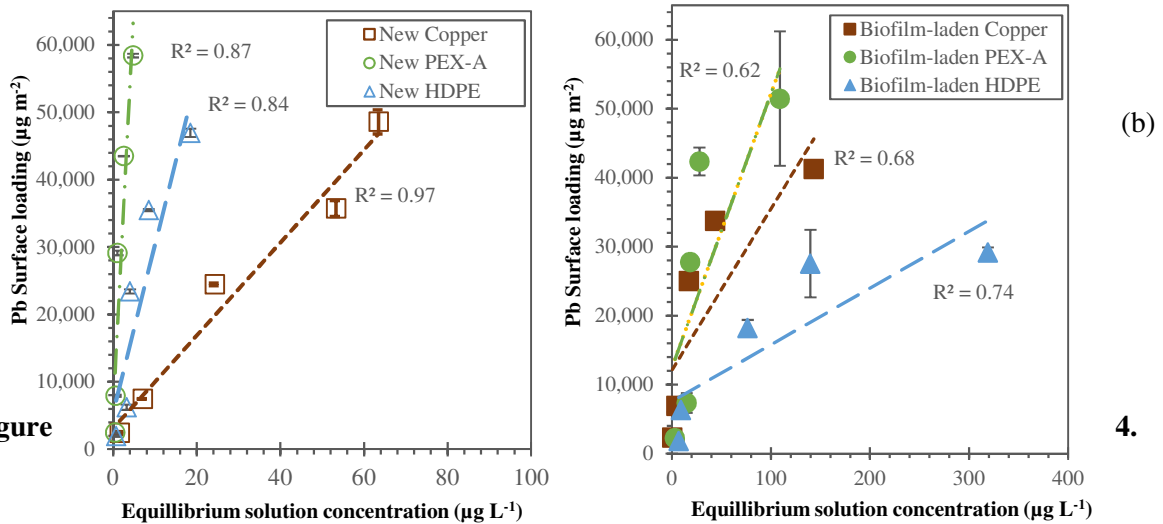
399 calculation of the Reynolds number ( $Re < 1$ ) for this system showed the laminar flow, some biofilm  
400 may have been detached from the pipe surface due to the alteration of water chemistry, biofilm  
401 maturation, and nutrient limitations. There was 18 h of flow and 6 h of stagnation during the biofilm  
402 growth process; however, the metal accumulation experiments for flow conditions were conducted for  
403 a continuous 48 h. Thus, this alteration of the water flow pattern may have contributed to the biofilm  
404 detachment; nutrient starvation and/or biofilm aging could reduce the biofilm matrix's internal  
405 cohesive strength and cause its detachment under lower shear stresses.

406 With increasing the Pb initial concentration, mostly Pb precipitates are present within the  
407 system. Thus, the surface sites available on both new and biofilm-laden water pipes act as the  
408 nucleation sites for the Pb precipitation and crystal growth as reported in our last study (Salehi et al.,  
409 2017). A similar trend of copper > PEX-A > HDPE was found for the Pb accumulation rates under  
410 stagnant conditions for both new and biofilm-laden water pipes. Thus, it could be inferred that copper  
411 provided more nucleation sites than both PEX-A and HDPE pipes. This finding is similar to the 5 d  
412 Pb exposure experiments, where the equilibrium Pb surface loading was greater for copper compared  
413 to the PEX-A and HDPE pipes. However, the Pb accumulation rates were not significantly different  
414 between PEX-A and HDPE pipes for both new and biofilm-laden pipes. Organics leached by plastic  
415 pipes may have made non-absorbable complexes with Pb and prevent their accumulation onto the  
416 pipe surface. Thus, a lower rate of Pb accumulation on plastic pipes was found compared to copper  
417 pipes. However, under the flow conditions, a greater volume of water (1 L) was in contact with the  
418 pipe segments thus, a lower concentration of the organics might be present within the system, which  
419 leads to forming of fewer complexes by Pb and consequently less prevention of its accumulation on  
420 the pipe surface. However, for both new and biofilm-laden water pipes, a greater rate of Pb  
421 accumulation was found for the PEX-A pipes compared to the HDPE pipes; it could be due to the  
422 smaller surface area of the PEX-A pipe segment ( $0.016 \text{ m}^2$ ) that was exposed to 1 L of Pb aqueous  
423 solution compared to the HDPE pipe segment ( $0.020 \text{ m}^2$ ).





424



427

**Figure**

**4.**

428 Equilibrium Pb surface loading for new and biofilm-laden PEX-A, HDPE, and copper pipes versus  
 429 residual Pb in the contact water under (a) stagnant and (b) flow conditions  
 430

431 Although the biofilm ( $2.97 \times 10^9$  copies  $\text{cm}^{-2}$ ) present onto the PEX-A pipes at the beginning  
 432 of the Pb exposure period was an order of magnitude higher than HDPE pipes ( $1.19 \times 10^8$  copies  $\text{cm}^{-2}$ ),  
 433 biofilm accumulation onto both plastic pipes resulted in a significant increase in Pb surface  
 434 loading. Biofilm accumulation onto the pipe surface could alter the metal accumulation behavior by  
 435 facilitating the biosorption and transport of metals across cell walls, complexation, precipitation, and  
 436 physical adsorption (Javanbakht et al., 2014). Moreover, as the zeta potential measurements revealed,  
 437 biofilm formation on the pipe surface could make the surface charge more negative and promote the  
 438 electrostatic attraction of the positively charged metal ions to the pipe surface.

439 Biofilm and heavy-metal interactions may occur through several potential mechanisms,  
 440 including microbial-mediated biosorption (Chen et al., 2022; Su et al., 2023), metal-mediated  
 441 alterations in microbial communities (Navarro-Noya et al., 2012; Caracciolo et al., 2021), metal-

442 mediated changes in biofilm gene expression (Portelinha & Angeles-Boza, 2021), and microbial-  
443 mediated metal transformations (Mohamed & Hatfield, 2005). Additional work is needed to identify  
444 the primary mechanism by which biofilms increase metal adsorption. Adsorption and sequestration of  
445 heavy metals into biofilms are facilitated by two major steps: 1) Metal ions undergo bulk diffusion  
446 into biofilm extracellular matrices, which is a complex amalgam of exopolysaccharides, proteins, fats,  
447 and free DNA that is heterogeneously and locally charged, and 2) Metal ions interact with solid  
448 surface and biofilm cellular walls. This interaction is complex and includes biosorption,  
449 bioaccumulation, resistance, and detoxification processes. Bioaccumulation occurs through metabolic  
450 activity between metals and living cells and may facilitate community turnover via local toxicity,  
451 favoring community members who can utilize or be minimally affected by these heavy metal species,  
452 which can lead to rapid turnover in biofilm communities when exposed to heavy metals (Ancion et  
453 al., 2010; Grün et al., 2018) **Figure SI-4** demonstrates the potential mechanisms of the heavy metal  
454 accumulation process onto the new and biofilm-laden plastic pipes (Galarce et al., 2020).

#### 455 **4. Conclusion**

456 In this study, the mechanistic role of biofilm presence on the Pb accumulation onto the PEX-A,  
457 HDPE, and copper potable water pipes was investigated under stagnant and flow conditions. The  
458 biofilm presence on the plastic pipes' inner walls promoted the negative surface charge and provided  
459 more available surface sites for Pb accumulation under stagnant conditions; however, biofilm  
460 presence on the copper pipes did not enhance the Pb uptake, which might be due to its saturation with  
461 the  $\text{Cu}^{2+}$  ions. The water flow conditions significantly influenced the extent of Pb accumulation onto  
462 the new and biofilm-laden water pipes. Although biofilm presence significantly increased the Pb  
463 accumulation onto the PEX-A and HDPE pipes, it did not alter the Pb accumulation onto the copper  
464 pipes, under stagnant conditions. Under flow conditions, Pb accumulation onto both biofilm-laden  
465 PEX-A and HDPE pipes was reduced compared to their new pipes, which might be due to the shorter  
466 interactions of Pb species with the biofilm surface. Under stagnant conditions, biofilm presence on the  
467 pipe surface enhanced the Pb accumulation by possibly facilitating the biosorption and transport of  
468 metals across cell walls, complexation, precipitation, and physical adsorption. This study will provide  
469 a foundation for continued exploration of heavy metal fate in water infrastructure and provide an  
470 understanding of metal adsorption and biofilm interactions. Additionally, understanding how biofilm  
471 affects the surface charge of the pipes could have broader implications beyond heavy metals, as it may  
472 influence the fate and transport of other contaminants, including organics and microbiological agents.

#### 473 **Acknowledgments**

474 Funding for this work was provided by the National Science Foundation (NSF) grant CBET-2029764.  
475 The authors thank Barry Wymore, the research technician in the College of Engineering at the  
476 University of Memphis, for building the pipe rig. The authors also thank Shima Ghoochani, the

477 graduate research assistant, and Colton Kirby, Carla Meier, Lauren Nichole Mitchell, and Rhianna  
478 Cameron Munns undergraduate research assistants in the Civil Engineering Department at the  
479 University of Memphis, for their assistance with conducting the metal exposure experiments. The  
480 authors would also like to thank Amy Abell and Nathan Mullins in the Department of Biological  
481 Sciences at the University of Memphis for their assistance in conducting ddPCR-based biomass  
482 quantification, Lingyun Peng in Clemson University for assistance with surface charge analysis, and  
483 Dibya Datta from University of Missouri for conducting the organic leaching experiments.

484

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